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AUSTRIAN STR POPULATION DATA USING THE POWERSEQ GY46 SYSTEM AND MASSIVELY PARALLEL SEQUENCING Petra Müller¹, Burkhard Berger¹, Martin Bodner¹, The DNASEQEX Consortium, Walther Parson^{1,2}



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Background: By using massively parallel sequencing (MPS) technologies to genotype forensic markers, it is possible to characterize sequence variations located in the repeat and flanking regions of short tandem repeats (STRs). To account for sequence based allele frequencies valid for the Austrian population, we performed a population study including 248 unrelated male donors verifiably born in Austria. In accordance with the Austrian law and permission of the Federal Ministry of the Interior, Austria, it is possible to use DNA samples and DNA profiles of the Austrian National DNA Database for population studies, preconditioned the data is made anonymous.

Concordance to capillary electrophoresis (CE) was assessed for all samples using the AmpFISTR NGM Select PCR Kit analysed on an Applied Biosystems Prism 3500XL Genetic Analyzer with the Gene Mapper ID-X software (all Thermo Fisher Scientific, Waltham, USA). MPS libraries were prepared manually using the PowerSeq GY46 system (Promega, Madison, USA) according to the manufacturer's recommendations. Sequencing was performed on the MiSeq FGx benchtop sequencer (Illumina, San Diego, USA) and analysed using STRait Razor v2 software [1].



Figure 2. We observed a 13 base pair deletion at **Penta D** (Chr21:43636191– 43636203, del-GAAAGAAAAAAAA), w results in allele 2.2 using length-based technologies.



Penta D exemplifies possible alignm challenges and discordances to Alignment was ambiguous due to proximity of the deletion to the rep region.

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| nent CE. | Length-b | ased | 45056072 | 45056074 | 45056075 | 45056076 | 45056077 | 45056078 | 45056080 | 45056081 | 45056082 | 45056083 | 45056084 45056085 | 45056086 | 45056087 | 45056088 | 45056089 | 45056090 | 45056091 | 45056092 45056092 | 45056094 | 45056095 | 45056096 | 46056097 | 45056098 | 45058099 | 45056101 | 46056102 | 45056103 | 45056104 | 45056105 | 45056106 | 46056107 46066107 | 45056109 | 45056110 | 45056111 | 45056112 | 45056113 | 45056114 | 45056115 46066116 | 45056117 | 46056118 | 45056119 | 45056120 | 45056121 | 45056123 | 45056124 | 45056125 |
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| peat | 7 | | G | A A | A A | G | А | A a | A A | A | Α | А | AG | A | A | Α | G | A | A | A A | A G | A | А | А | A | G | A A | A A | A | G | А | А | A / | A G | A | A | Α | А | G | A A | A A | A | G | А | | | | |
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Figure 3. Stutter analysis was performed on a subset of 50 samples (corresponding to 20% of the entire sample set). Samples were selected according to the total numbers of reads (selection criteria: $\leq 63,300$ reads or $\geq 199,000$ reads). a) Violin plot of relative stutter ratios for 22 aSTRs included in the PowerSeq GY46 panel. Median stutter ratios ranged from 1.9% (PentaD) to 14.7% (D22S1045). b) Locus specific violin plot for D1S1656 showing distinct differences in stutter ratios between integer alleles and micro-variants (global median: 13.1%, median stutter ratios ranged from 8.3% to 22.3%).

a) ₃₀₁

b)

Conclusions:

In general, the here presented results underline the preliminary significance of sequence based studies. Our results were 99.99% concordant to known genotypes (derived from CEanalysis) including 15 aSTRs (AmpFISTR NGM Select PCR Kit). Analysing 248 samples, MPS was able to successfully

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References: [1] King JL, Wendt FR, Sun J, Budowle B. STRait Razor v2s: Advancing sequence-based STR allele reporting and beyond to other marker systems. Forensic Sci Int Genet. 2017;29:21-8. [2] Devesse L, Ballard D, Davenport L, Riethorst I, Mason-Buck G, Syndercombe Court D. Concordance of the ForenSeq system and characterisation of sequence-specific autosomal STR alleles across two major population groups. Forensic Sci Int Genet. 2017;34:57-61. [3] Gettings KB, Borsuk LA, Steffen CR, Kiesler KM, Vallone PM. Sequence-based U.S. population data for 27 autosomal STR loci. Forensic Sci Int Genet. 2018;37:106-15.

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